

# How to produce comparable data in conservation

# genetics for the Apennine brown bear

Scarpulla E.<sup>1</sup>, Antonucci A.<sup>2</sup>, Boattini A.<sup>1</sup>, Latini R.<sup>3</sup>, Mucci N.<sup>4</sup>, Pizzol I.<sup>5</sup>, Davoli F.<sup>4</sup>

<sup>1</sup> Department of Biological, Geological and Environmental Sciences, University of Bologna, Bologna, Italy. erminia.scarpulla@gmail.com <sup>2</sup> Ente Parco Nazionale della Majella, Uffici "Tutela Valori Naturali e Ambientali" e "Monitoraggio e Gestione Biodiversità", Sede Operativa - Badia Morronese, Sulmona (AQ), Italy. <sup>3</sup> Ente Autonomo Parco Nazionale d'Abruzzo, Lazio e Molise, Servizio Scientifico, Pescasseroli (AQ), Italy.

<sup>4</sup> Unit for Conservation Genetics (BIO-CGE), Department for the Monitoring and Protection of the Environment and for Biodiversity Conservation, Italian Institute for Environmental Protection and Research (ISPRA), Ozzano dell'Emilia, Italy.

<sup>5</sup> Regione Lazio, Direzione Capitale Naturale, Parchi e Aree Protette, Area Tutela e Valorizzazione dei Paesaggi Naturali e della Geodiversità.

## INTRODUCTION

The Apennine brown bear (U. arctos marsicanus) presents a low level of variability [1], therefore ISPRA conducted the individual identification on the basis of 11 Ursidae specific markers (STR) plus sex. In the last decade, two different labs (WGI, Wildlife Genetics International, B.C., Canada and ISPRA) conducted the genotyping of the Apennine brown bear. WGI added two markers designed on the domestic dog genome (CXX20 and REN144A06) and removed two ones that had been previously used. Thus, their total selection was of 11 markers, 9 of which in common with ISPRA (G1D, G10B, G10C, G10L, Mu05, Mu11, Mu50, Mu51, Mu59), with an additional marker in common to both labs for equivocal cases (G10P) [2]. For a population with a low variability it is important to select the optimal STR marker set for individual identification, in order to allow the correct identification of the individuals overtime and to reduce genotyping errors.

### **MATERIALS AND METHODS**

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ows an improvement, albeit not significant, of the discrimination capacity using the complete set of 13 STRs + AMG compared to the other STRs marker sets. However canid loci show a higher occurrence of genotyping errors.

Allelic patterns (Fig. 7) show slight variation over time and PCA (Fig. 6) shows a substantial overlap of genetic diversity in the two considered periods.





9.0E-03



0.540

0.520

0.500

0.480

0.460

0.440

0.420

0.400

Locus	Range (bp)	Conversion	<b>P</b> <sub>ID</sub>	<b>P</b> <sub>IDsib</sub>
CXX20	132-136	-3	0.22	0.50
REN144A06	110-130	+1	0.24	0.51
G1D	100-114	-72	0.25	0.53
Mu51	114-122	-92	0.26	0.53
G10B	112-128	-28	0.36	0.58
G10C	95-105	-102	0.37	0.59
Mu59	101-107	-128	0.38	0.60
Mu11	88-96	-100	0.39	0.62
Mu05	135-137	/	0.40	0.62
G10L	148-154	-9	0.41	0.63
Mu50	100-104	-32	0.41	0.63
G10P	152-164	+7	0.65	0.81
Mu15	117-121	Not used by	0.71	0.84
		WGI		
Amelogenin	158-212	-46/-38	-	-





parco



The following software were used for data analysis: • GenAlEx 6.4 for allelic patterns, H<sub>o</sub>, H<sub>e</sub>, HWE, P<sub>ID</sub>, P<sub>IDsib</sub>, number of MM.

• GIMLET 1.3.3 and MicroChecker 2.2.3 to estimate genotyping errors frequencies (ADO, FA, PCR+ and null alleles).

• R (*chisq.test* and *fisher.test*) for statistical significance among groups.

#### 0.14 8.0E-03 He FA PIDsib 7.0E-03 0.12 6.0E-03 0.1 5.0E-03 0.08 4.0E-03 0.06 3.0E-03 0.04 2.0E-03 0.02 1.0E-03 0.0E+00 GIOS GIOC GIOL GIOS GIO WAR WIT WIT WITH WAR WAR WITH STRs 11 STRs STRs of 13 STRs Fig. 5 - Genotyping errors at single locus. Fig. 3 - Mean H<sub>e</sub> and H<sub>o</sub> in the Fig. 4 - P<sub>ID</sub> and P<sub>ID</sub> sib in the four STR marker sets. four STR marker sets. Fig. 7 -**Principal Coordinates** Na 3.000 0.600 Fig. 6 - PCA Variation Na Freq 2.500 0.500 of the of allelic 0.400 sit >= 5% 2.000 Ne variability patterns **b** 1.500 1.000 0.300 8 from from 0.200 **t** 2000-2010 2000-2010 No. Private pre-0.500 0.100 Alleles to 2011to 2011arctos arctos& 0.000 0.000 2017. 2017. post arctos&post pre-arctos Coord. 1

PID

### CONCLUSIONS

• In order to avoid both underestimation (high values of P<sub>ID</sub>) and overestimation (high levels of ADO and FA) in genotyping results, future monitoring will be conducted using the ISPRA set of 11 Ursidaespecific STRs with the addition of CXX20, that minimize the risk of shadow effect (P<sub>ID</sub> = 8.6 \* 10<sup>-6</sup>; P<sub>IDsib</sub> = 3.0 \* 10<sup>-3</sup>). In addition, marker REN144A06 will be used to improve the discriminatory capacity in uncertain cases.

• The population shows a slight and not significant loss of diversity due to genetic drift (Fig. 6, Fig. 7). Therefore the chosen STR panel is suitable for individual identification in the near future, but markers with higher discriminatory power will be needed for parentage analysis (eg. panel of SNPs).

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